

Prevalence of *Haplorchis taichui* Infection in Snails from MaeTaeng Basin, Chiang Mai Province, by Using Morphological and Molecular Techniques

Thapana Chontanarth* and Chalobol Wongsawad*

Abstract

The investigation of biological diversity of intestinal trematode in snails which the report concerned is rarely and not extended in current status. Especially in minute intestine trematode, *Haplorchis taichui* Witenberg, 1930, family of Heterophyidae, which was found in the small intestine of mammals including human, and causes serious clinical problem worldwide. So, this study was aimed to investigate *H. taichui* infection in freshwater snails from Mae Taeng basin, Chiang Mai, Thailand. Total of 1,836 snails were collected during April 2008 to August 2012. Cercarial infection was examined by crushing method. Molecular identification of *H. taichui* were conducted by a DNA specific primer which amplified the mCOI gene. The PCR product of mCOI were sequenced and confirmed by BLAST program. Six types of cercariae were found viz. megalurous, furcocercous, monostome, pleurolophocercous, parapleurolophocercous, and virgulate cercaria. The parapleurolophocercous cercaria in *Melanoides tuberculata*, *Tarebia granifera*, and *Thiara scabra* were larval stages of *H. taichui* which yield the specific fragment of 256 bp. Overall of *H. taichui* infection of snails was 42.71%. The mCOI sequences had 99% similarity with the 29 isolated gene references of the *H. taichui* in Genbank data base. The molecular method had suitability as an epidemiological tool for suitable control programs against the dissemination of trematodes.

Keywords : Intestinal trematodes Prevalence Molecular identification mCOI gene

* Department of Biology Faculty of Science Chiang Mai University Chiang Mai 50202 Thailand.



ค่าความชุกของการติดพยาธิใบไม้ชนิด *Haplorchis taichui* ในหอยเดียวจากลุ่มแม่น้ำแม่แตง จังหวัดเชียงใหม่ โดยเทคนิคทางสัณฐานวิทยาและอนุชีววิทยา

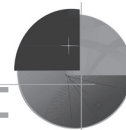
ฐาปนา ชลธนานารณ* และ ชโลบล วงศ์สวัสดิ์*

บทคัดย่อ

การศึกษาความหลากหลายทางชีวภาพของพยาธิใบไม้ในลำไส้ (intestinal trematode) มีรายงานน้อยมากและยังไม่ครอบคลุม โดยเฉพาะพยาธิใบไม้ในลำไส้ขนาดเล็ก *Haplorchis taichui* Witenberg, 1930 ในวงศ์ Heterophyidae เป็นปรสิตในสัตว์เลี้ยงลูกด้วยนมรวมถึงมนุษย์ การติดเชื้อพยาธิใบไม้ก่อให้เกิดปัญหาที่สำคัญทางสาธารณสุข และมีรายงานการระบาดในประเทศไทยอย่างต่อเนื่อง ดังนั้นการศึกษานี้มีวัตถุประสงค์เพื่อศึกษาการติดเชื้อพยาธิใบไม้ *H. taichui* ในหอยฝาดเดียวจากลุ่มแม่น้ำแม่แตงจังหวัดเชียงใหม่ โดยทำการเก็บหอยฝาดเดียวทั้งหมด 1,836 ตัว ระหว่างเดือนเมษายน 2008 ถึง เดือนสิงหาคม 2012 เพื่อตรวจสอบการติดตัวอ่อนพยาธิใบไม้โดยวิธี crushing สำหรับการระบุอัตลักษณ์ของพยาธิใบไม้ชนิด *H. taichui* จะถูกศึกษาโดยเทคนิคทางอนุชีววิทยาด้วยไพรเมอร์แบบจำเพาะและถูกเพิ่มปริมาณดีเอ็นเอของยีน mCOI (mitochondrial cytochrome c oxidase subunit I) เพื่อหาลำดับเบสและตรวจสอบด้วยโปรแกรม BLAST จากผลการศึกษาพบเชอร์คาเรียทั้งหมด 6 รูปแบบ คือ megalurous cercaria furcocercous cercaria monostome cercaria pleurolophocercous cercaria parapleurolophocercous cercaria และ virgulate cercaria การตรวจสอบการติดพยาธิ *H. taichui* ระยะเชอร์คาเรียในหอยโดยเทคนิคทางอนุชีววิทยาพบว่าเชอร์คาเรียรูปแบบ parapleurolophocercous cercaria ให้ผลการทดสอบเป็น positive result โดยสามารถเกิดแถบดีเอ็นเอขนาด 256 bp ที่มีความจำเพาะต่อพยาธิ *H. taichui* มีค่าความชุกรวมร้อยละ 42.71 ส่วนลำดับเบสของยีน mCOI มีความคล้ายคลึงเท่ากับร้อยละ 99 จาก 29 ไอโซเลทของ *H. taichui* ในฐานข้อมูลของ Genbank การศึกษาในครั้งนี้ชี้ให้เห็นว่าเทคนิคทางอนุชีววิทยาสามารถนำมาประยุกต์เป็นเครื่องมือในการศึกษาด้านระบาดวิทยาเพื่อใช้ในการวางแผนควบคุมการกระจายตัวของพยาธิใบไม้ได้อย่างมีประสิทธิภาพยิ่งขึ้น

คำสำคัญ : พยาธิใบไม้ ค่าความชุก การระบุชนิดเชิงโมเลกุล ยีนไซโตโครมซี ออกซิเดส

* ภาควิชาชีววิทยา คณะวิทยาศาสตร์ มหาวิทยาลัยเชียงใหม่ จังหวัดเชียงใหม่ 50200



Introduction

Mae Taeng basin is located in Chiang Mai province, northern Thailand (Figure 1). It serves as an important area for both aquatic culture and fisheries. Much of this area is in Mae Ngad Somboonchon Dam irrigating channels which have high biodiversity of aquatic animals that are associated in the completion of trematode life cycles (1, 2). There have been several reports the heterophyid trematode infections in freshwater animals in Chiang Mai (3), especially, the minute intestinal trematode, *Haplorchis taichui*, Witenberg, 1930 which have a high prevalence of infection in cyprinoid fishes (4-6).

The life cycle of *H. taichui*, involves freshwater snails as the first intermediate host, cyprinoid fishes as second host, and with

fish-eating mammals as definitive hosts (7, 8).

The pleurolophocercous and parapleurolophocercous cercariae have been reported heterophyid trematodes including *H. taichui*, *H. pumilio*, *Stellantchasmus falcatus*, and *Centrocestus caninus*, which are abundantly recovered from freshwater snails in northern Thailand (9-11).

The traditional method of *H. taichui* detection in snails is usually performed by exposing the snails to light or by dissection, in which cercariae may be observed. These methods are difficult to detect specific parasite stages to the species because of similar morphological traits in closely related species the Heterophyidae. An accurate identification of cercarial stage is very important for understanding the epidemiological situation

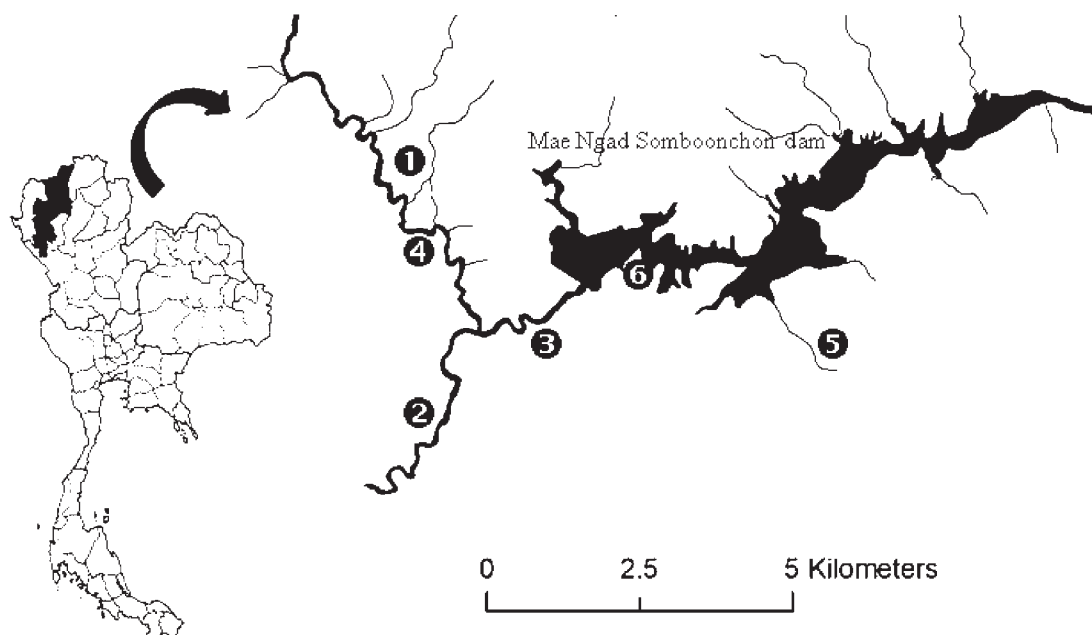
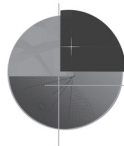


Figure 1 A map of Thailand showing the collecting sites in Mae Taeng basin, Chiang Mai province, Thailand. (1) irrigation canal (2-3) rice paddys (4-5) Mae Taeng River (6) Mae Ngad Somboonchon Dam.



and for developing effective control measures.

Various PCR methods have been developed for detection of trematode species in intramolluscan stages namely, Amplified Fragment Length Polymorphism (AFLP), High Annealing Temperature – Random Amplified Polymorphic DNA (HAT-RAPD), Polymerase Chain Reaction – Restriction Fragment Length Polymorphism (PCR-RFLP), and gene sequencings (12-14). The development of aspecific DNA primer for *H. taichui* detection has been rarely successful. Some report have been established specific primers for the detection of *H. taichui* by generating the 256 bp specific fragment as a positive result (6).

Data from mitochondrial-based PCR have been extensively used in studies of identification, evolution, phylogeny, biogeography, population genetics and systematic because the mitochondrial genomes are readily abundantly copied and not strikingly different in related species (15). Lee et al. compared the phylogenetic relationship of these heterophyid trematodes, viz. *Metagonimus yokogawai*, *M. takahashii*, and *M. miyatai*, using 28S rDNA and cytochrome c oxidase subunit I (mCOI) gene. The results showed that the phylogenetic tree from 28S ribosomal DNA was similar with mCOI sequence data (16).

Our study provides data on the prevalence of *H. taichui* infections in freshwater snails from Mae Taeng basin, Chiang Mai, Thailand by morphological and PCR-based methods. This is the first step in

providing new data on the recent distribution and prevalence of trematode infections and for developing effective control measures.

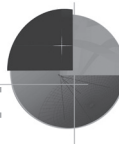
Method

Study sites

Six sites were selected based on areas with public health problems with trematode infections in and around the Mae Ngad Somboonchon Dam, Mae Taeng district, Chiang Mai province. These areas include an irrigation canal (N 21°/12.864/ E 0.50°/08.488/), rice paddies (N 21°/13.134/ E 0.50°/07.982/ and N2116.486 E 0.409.359), Mae Taeng River (N 21°/12.992/ E 0.50°/07.862/ and N2112.992 E 0.5007.862), and the dam (N2119.725 E 0.5003.645) (Figure 1).

Snail samples and cercarial infections

Snail specimens were collected by hand during April 2008 to June 2010. After collection, the snails were identified by using Brandt (17) and were examined for cercarial infection using a crushing method. Each groups of cercaria were stained in Delafield's haematoxylin, dehydrated in an alcohol series, cleared with xylene, and mounted in permount. The cercariae were then identified to family level and in some cases to genus (18, 19). The pleurolophocercous and parapleurolophocercous cercariae were preserved in 70% alcohol for DNA extraction since these cercarial types have been reported as minute intestinal heterophyid trematodes (20).



Genomic DNA extraction and PCR amplification

The genomic DNA was extracted and purified individually from both cercarial types found in each snail using a commercial GF-1 tissue DNA extraction kit (Vivantis, Malaysia) according to the manufacturer's instructions.

Molecular identification by DNA specific primer

PCR was performed to detect the *H. taichui* infections in snails. The protocol followed Wondsawad et al. (6). Sequences of the primers used in this PCR amplification were Forward 5'-GGCCAACGCAATCGTCATCC-3' and Reverse 5'-GCGTCGGGTTTCAGACA TGG-3'. Using these primers, genomic DNA was amplified in 20 µl final volume, including 50 ng of DNA template, 50 pM/ l of each primer (forward and reverse), 1.5 mM of MgCl₂, 200 µ of dNTPs and 0.5 unit of Taq DNA polymerase, and volume made to 20 µl with deionized water.

Amplification was perform in a thermocycles with the initial denaturation step at 95°C for 5 minutes, then 35 cycles including denaturation at 95°C for 45 seconds, primer annealing at 68°C for 45 seconds, extension at 72°C for 1 minutes, and final extension at 72°C for 7 minutes. PCR products were analyzed through 1.8% agarose gels electrophoresis separation at 50 volts for 40 minutes in TBE buffer, stained with ethidium bromide, and visualized with a Kodak digital camera (Gel Logic 100).

The genomic DNA of positive results, which gave an amplification of the 256 bp

fragment, were amplified the barcoding region mCOI gene by a pair of primers according to Yu et al. (21). It included (JB3) 5' TTTTGGGCATCC TGACGTTTAT 3', as a forward primer, and (JB 4,5) 5' TAAAGAAAGAACATAATGAAAATG 3', as a reverse primer.

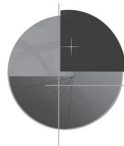
The specific DNA fragment of mCOI were direct sequenced. And confirmed by BLAST (Basic Local Alignment Search Tool) program in the NCBI (National Center for Biotechnology Information).

Results

Prevalence of infection

1,836 snails of 11 species were collected, viz. *Bithynia siamensis*, *Clea helena*, *Eyriesia eyriesi*, *Filopaludina doliaris*, *F. martensi*, *F. sumatrensis*, *Lymnaea auricularia*, *Melanoides tuberculata*, *Pila polita*, *Tarebia granifera*, and *Thiara scabra*. The results showed that 315 out of 1,836 snails had cercarial infection, with an overall prevalence of 17.16% (Table 1). *M. tuberculata*, *L. auricularia*, and *T. granifera* were the most infected and susceptible hosts for cercarial infections with a prevalence of 35.71%, 33.33%, and 30.50%, respectively. *C. helena*, *F. sumatrensis*, *F. doliaris*, and *P. polita* were not infected.

Six groups of cercariae were detected: megalurous, furcocercous, monostome, pleurolophocercous, parapleurolophocercous, and virgulate cercaria. They were subdivided into 7 types based on morphological characteristics and internal organ arrangement. They are described here (Figure 2).

**Table 1** Prevalence of cercarial infections in snails from the Mae-Taeng basin, Chiang Mai, Thailand

Snail species	Snail samples	Infected samples	Prevalence (%)	Type of cercariae	Prevalence of cercarial types (%)
Family Bithyniidae					
<i>Bithynia siamensis</i>	150	16	10.67	Notocotylidae	7.33
				<i>Haplorchis</i> sp.	1.33
				Strigeidae	0.68
				Lecithodendriidae	1.33
Family Buccinidae					
<i>Clea helena</i>	2	0	0.00	-	-
Family Viviparidae					
<i>Eyriesia eyriesi</i>	80	2	2.50	<i>Haplorchis</i> sp.	2.50
<i>Filopaludina doliaris</i>	150	0	0.00	-	-
<i>Filopaludina martensi</i>	296	2	0.68	Strigeidae	0.34
				Lecithodendriidae	0.34
<i>Filopaludina sumatrensis</i>	120	0	0.00	-	-
Family Lymnaeidae					
<i>Lymnaea auricularia</i>	30	3	33.33	Heterophyidae	33.33
Family Thiaridae					
<i>Melanoides tuberculata</i>	448	160	35.71	Distome	2.01
				Lecithodendriidae	3.12
				<i>Haplorchis</i> sp.	30.56
				Strigeidae	0.22
				Heterophyidae	8.48
<i>Tarebia granifera</i>	377	125	33.16	Philophthalmidae	0.53
				<i>Haplorchis</i> sp.	28.64
				Lecithodendriidae	0.53
				Notocotylidae	0.16
				Heterophyidae	0.82
<i>Thiara scabra</i>	93	7	7.22	Heterophyidae	5.38
				Lecithodendriidae	1.08
				<i>Transversotrema</i> sp.	1.08
				<i>Haplorchis</i> sp.	4.30
Family Ampullariidae					
<i>Pila polita</i>	90	0	0.00	-	-
Total	1,836	315	17.16		

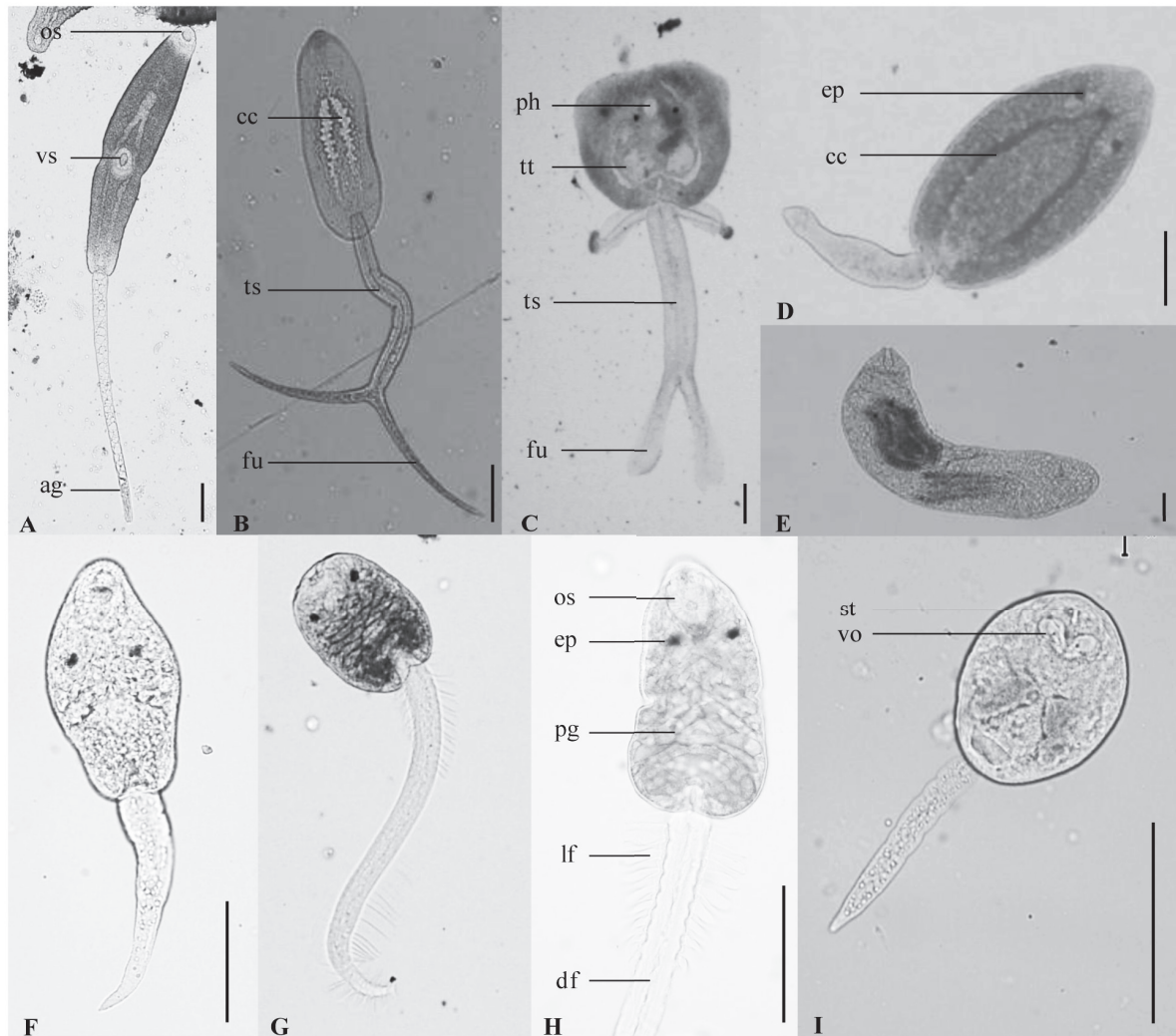
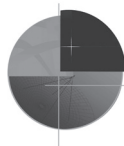


Figure 2 Seven types of cercaria were found infect to snails. **A** – distome cercaria: Philophthalmidae cercaria, **B** – furcocercous cercaria, Strigeidae cercaria, **C** – furcocercous cercaria: *Transversotrema* sp., **D** – monostome cercaria: Notocotylidae cercaria, **E** - redia of Notocotylidae, **F** – pleurolophocercous cercaria, Heterophyidae cercaria, **G** – parapleurolophocercous cercaria: *Haplorchis* sp. cercaria (body), **H** - anterior part of *Haplorchis* sp., **I** –virgulate cercaria: Lecithodendriidae cercaria (ag - adhesive gland, df - dorsoventral nfold, ep - eye spots, fu - furca, lf - lateral nfold, os - oral sucker, pg - penetrated gland, ph - pharynx, st - stylet, ts – tail stem, tt – testis, vo - virgulate organ, vs – ventral sucker) (scale bar 0.1 mm).

**Megalurous cercaria: Philophthalmidae**

This parasite only infected *T. granifera*. The megalurous cercaria is ovoid with yellowish-brown granules. The body surface is covered with longitudinal and fine traverse wrinkles. Minute spines cover the surface of the posterior half of the body. The esophagus bifurcated midway between the pharynx and the ventral sucker, and the intestinal ends blindly near the posterior end of body. The ventral sucker is slightly larger than the oral sucker, and is located medially on the the body. The tail is as long as the body, elastic, and slender. Long tail tips numerous adhesive gland cells and lacks an excretory canal in the tail base (Figure 2A).

Furcocercous cercaria: Strigeidae

This parasite was found in *B. siamensis*, *F. martensi*, and *M. tuberculata*. The cercaria body is ovoid and the body surface is covered with minute spines. The oral sucker is subterminal. The pharynx is very small and circular. The large intestine is composed of two ceca. The tail is forked and longer than the body. The tail stem is longer than the furca, and each furca has a dorsal and ventral finfold (Figure 2B).

Furcocercous cercaria: Transversotrema sp.

Transversotrema cercaria was found only in *T. scabra*. The cercaria body is short, flat, discord, and with yellowish-brown pigmentation. The body surface is covered with scale-like spines. The pharynx is very large and the esophagus is short and narrow. There are one pair of eyespots which are large spherical, located posterior to the pharynx.

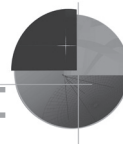
An oral sucker is absent while a ventral sucker is present medially. Testes are symmetrical and within the intestinal ring, while the genital pore is located medially. The tail is longer than the body and with arm like processes at the anterior end of the tail-stem (Figure 2C).

Monostome cercaria: Notocotylidae

Notocotylidae cercariae were found only in *B. siamensis* and *T. granifera*. The cercaria body is ovoid and the anterior quarter of the body is highly pigmented, especially around the eyespots. There are three black pigmented eyespots arranged in the form of a triangle in the anterior dorsal region of the mature cercaria (Figure 2D).

Pleurolophocercous cercaria: Heterophyidae

This type of cercaria was found in *B. siamensis*, *L. auricularia*, *M. tuberculata*, *T. granifera*, and *T. scabra*. The cercaria is ovoid and entirely covered with minute spines. A pair of conspicuous eyespots rectangular are found at the end of the anterior third of the body. The pharynx lies just behind the oral sucker. Intestinal ceca bifurcate and extend posteriorly to the excretory vesicle level. Seven pairs of penetration glands are arranged in 2 bundles anterolaterally to the genital premordium, which is a relatively large triangular mass of cells located just anterior to the excretory vesicle. The tail is almost the as long as body, slender, and usually attached to the dorso-ventral finfolds. A brownish pigment is dispersed throughout the body except in the oral sucker (Figure 2F).



Parapleurolophocercous cercaria : *Haplorchis* sp.

This type of cercaria was found in *B. siamensis*, *E. eyriesi*, *M. tuberculata*, *T. granifera*, and *T. scabra*. The cercaria is ovoid and is entirely covered with minute spines. The small pharynx lies just behind the oral sucker. Two square eyespots are located on each side above the pharynx. The anterior part of the body is covered with fine spines. Seven pairs of penetration glands are located between the pharynx and the posterior part of the body, surrounding mainly the genital premordium and arranged into two columns with the ventral sucker. The excretory vesicles are elongate. The tail is slender about 3 times the body length with two lateral finfolds extending along the anterior third of the tail. A dorsoventral finfolds extends along the posterior two-thirds and around tip of the tail (Figure 2 G).

Virgulate cercaria: Lecithodendriidae

This cercaria was found in *B. siamensis*, *F. martensi*, *M. tuberculata*, *T. granifera*, and *T. scabra*. The cercaria is small shaped with a prominent stylet and develops in sporocysts.

The oral sucker extends to a pharynx; oesophagus bifurcating preacetabularly into an intestinal caeca which encircles the acetabulum, terminating postacetabularly. The acetabulum is smaller than the oral sucker. The virgulate organ is butterfly-shaped, situated below the stylet. There are three pairs of penetration glands, the two anterior pairs have fine granules and the posterior pair with coarser granules medially. The tail is shorter than the body (Figure 2I).

H. taichui infection in snails

H. taichui specific primers were performed to amplify a 256 bp fragment in the PCR technique for the identification of *H.taichui* cercariae infecting snails. Amplification of *H. taichui* cercariae could detect only the 256 bp specific fragment. Genomic DNA of adults from this parasite were used as a positive control, which also gave 256 bp specific fragment.

As shown in Table 2, 199 of 1,836 snails were infected by parapleurolophocercous cercaria including *B. siamensis*, *E. eyriesi*, *M. tuberculata*, *T. granifera*, and *T. scabra*.

Table 2 Prevalence of *H. taichui* infections, using specific PCR.

Snail species	Number infected with parapleurolocercouse cercaria	Positive	Prevalence (%)
<i>Bithynia siamensis</i>	2	0	0.00
<i>Eyriesia eyriesi</i>	2	0	0.00
<i>Melanoides tuberculata</i>	98	42	42.86
<i>Tarebia granifera</i>	93	42	45.16
<i>Thiara. scabra</i>	4	1	25.00
Total	199	85	42.71

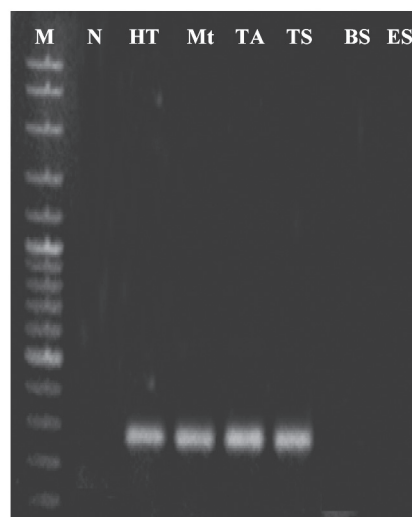
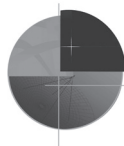


Figure 3 Gel electrophoresis proles of parapleurolophocercous cercariae isolated from thiarid snails. Lane M: molecular size maker, Lane N: negative control, Lane Ht: adult of *H. taichui*, DNA samples of parapleurolophocercous cercariae form snails (Lane Mt: *Melanoides tuberculata*, Lane Ta: *Tarebia granifera*, Land Ts : *Thiara scabra*, Lane Bs: *Bithynia siamensis*; Lane Ee: *Eyriesia eyriesi*).

```

1AGTGAGACACATTTGTAGGACGTTAACAAATAAAGATTCCTTGTTTGGTT
  ATGGGGGTTTAGTTCTTGCTATGTTTTCTATAGTCTGTTTGGGGAGTGTT
  GTTTGAGCTCATCATATGTTTATGGTTGGGTTGGATGTTAAGACGGCTGT
  TTTTTTAGTTCTGTGACTATGGTTATAGGAGTCCCCACAGGTATAAAGG
  TTTTTCTTGGCTGTATATGTTGGCGGGAAGTCGGGGCCGGTTTTGGGAT
  CCGATAATGTGGTGGATATTGGGTTTTATTGTCCTATTTACTATCGGGGG
  GGTGACCGGGATTGTGTTGTCTTCTCCTATAATGGATA           340

```

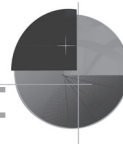
Figure 4 Sequence of mCOI nucleotides of *Haplorchis taichui*.

After testing with *H. taichui* specific primers, only three species were infected this trematode, viz. *M. tuberculata*, *T. granifera*, and *T. scabra* (Figure 3). The overall prevalence of *H. taichui* infections based on PCR detection was 42.71%. *T. granifera* was the most susceptible snail host for *H. taichui* infections (Table 2).

The length of partial mCOI nucleotide sequence data was 340-bp. The mCOI sequence showed 99% similarity with the reference sequence of the 29 isolated *H. taichui* in the Genbank data base (Figure 4).

Discussion

Eleven snail species from the Mae Taeng basin are intermediate hosts of digenetic trematodes. We found, 7 cercarial types in 4 snail families, viz. Thiaridae, Viviparidae, Bithyniidae, and Lymnaeidae) from 7 snail species (Table 1). The prevalence of cercarial infections found in our study (17.16%) was higher than Mard-arhin et al. found in five provinces in northern Thailand (6.20 %) (22). Dechruruxsa et al. reported that the prevalence of cercarial infections in thiarid



snails from the Khea River, Phitsanulok province, Thailand was only 0.9% (11). The *Transversotrema* sp. cercaria infected only *T. scabra* which agrees with Wongsawad and Kumchoo (23).

In our study, parapleurolophocercous and pleurolophocercous cercariae were dominant. They have been reported previously as heterophyid trematodes, viz. *C. caninus*, *H. taichui*, *H. pumilio*, and *S. falcatus* (11, 13, 24). Our findings agree with Radomyos et al. who reported that heterophyid trematodes are widely distributed and with high prevalence in northern Thailand (25). Kumchoo et al. reported a high prevalence of *H. taichui* infections (91.4%) in cyprinoid fish from Mae Taeng district, Chiang Mai province (5). Nithikathkul and Wongsawad (2008) indicated a high prevalence of *H. taichui* and *Haplorchoides* sp. infections (83.90%) in Chiang Mai province (26).

For the PCR-based assay, parapleurolophocercous cercariae were confirmed to be *H. taichui*. Overall *H. taichui* infections based on specific PCR amplification was 42.71% of all parapleurolophocercous cercariae populations. The prevalence of the morphological method was higher than the molecular method because the PCR-based method is more exact specific than the identification by using only morphology. Pleurolophocercous and parapleurolophocercous cercariae can develop to be several heterophyid species. Our study provides significant evidence for the identification and epidemiological situation of larval trematodes in their intermediate hosts. The high similarity of mCOI with 29 isolates of *H. taichui* from Vietnam and Thailand

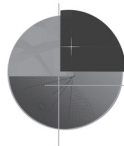
indicated that the mCOI sequence products good to species level and applicable for molecular identification. According to Thaekham et al. investigated the PCR-RFLP patterns of cytochrome c oxidase subunit I (mCOI) gene amplification to distinguish *O. viverrini* from *H. taichui* (12).

Our results established that three species of snails (*M. tuberculata*, *T. granifera*, and *T. scabra*) serve as the first intermediate hosts of *H. taichui*. Dechruksa et al. reported that thiarid snails were infected with intestinal and blood fluke larvae (11). The snails, *M. tuberculata* and *T. granifera* are medically important since they can serve as the first intermediate host of minute intestinal flukes (13, 24).

The PCR-based method is a more effective approach for the identification of *H. taichui* cercariae including other medically important species, which can be used for epidemiological studies, and developing a control program.

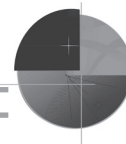
Acknowledgements

We greatly appreciate the Applied Parasitology Research Laboratory, Institute for Science and Technology Research and Economic Plants Genome Research and Service Center, Chiang Mai University, Thailand for providing research facilities. Special thanks are given to Mr. Sukson Chuboon for his helpful and suggestions. We would like to thank Dr. J.F. Maxwell for editing our manuscript. Finally, we acknowledge the graduate school, Chiang Mai University for supporting the publication fee of this paper.



References

1. Jousson, O., Bartoli, P. Pawloski J. : Molecular identification of developmental stages in opecoelidae (Digenea). Int Parasitol. 29:1835-58, 1999.
2. Sri-aroon, P., Butraporn, P., Limsomboon J. et al. : Freshwater mollusks of medical importance in Kalasin province, northeast Thailand. Southeast Asian J Trop Med Public Health 34(3):653-7, 2005.
3. Boonchot, K., Wongsawad, C. : A survey of helminths in cyprinoid fish from the Mae Ngad Somboonchon reservoir, Chiang Mai Province, Thailand. Southeast Asian J Trop Med Public Health 36(1):103-7, 2005.
4. Chai, J.Y., Lee, S.Y. : Food-borne intestinal trematode infections in the republic of Korea. Parasitol International 51(1):129-54, 2002.
5. Kumchoo, K., Wongsawad, C., Chai J.Y., et al. : High Prevalence of *Haplorchis taichui* Metacercariae in cyprinoid fish from Chiang Mai Province, Thailand. Southeast Asian J Trop Med Public Health 36(2):451-5, 2005.
6. Wongsawad, C., Wongsawad, P., Chai J.Y. et al. : *Haplorchis taichui*, Witenberg, 1930: development of a HAT-RAPD marker for the detection of minute intestinal fluke infection. Exp Parasitol 123(1):158-61, 2009.
7. Dzikovski, R., Levy, M.G., Poore, M.F., et al. Use of rDNA polymorphism for identification of Heterophyidae infecting freshwater fishes. Dis Aquat Org 59(1):35-41, 2004.
8. Watthanakulpanich, D., Waikagul, J., Maipanich, W., et al. : *Haplorchis taichui* as a possible etiologic agent of irritable bowel syndrome-like symptoms. Korean J Parasitol. 48(3):225-9, 2010.
9. Umadevi, K., Madhavi, R. : Observations on the morphology and life-cycle of *Procerovum varium* (Onji & Nishio, 1916) (Trematode: Heterophyidae). Sys Parasitol. 46(1):215-25, 2000.
10. Boga, T., Corddeiro, F.M., Gouveia, J.S. : *Melanoides tuberculata* (Gastropoda: Thiaridae) as intermediate host of Heterophyidae (Trematoda: Digenea) in Rio de Janeiro Metropolitan area, Brazil. Rev Inst Med Trop S Paulo 47(2):87-90, 2005.
11. Dechruksa, W., Krailas, D., Ukong, S., et al. : Trematode infections of the freshwater snail family Thiaridae in the Khek River, Thailand. Southeast Asian J Trop Med Public Health 38(6):1016-28, 2007.
12. Thaenkharn, U., Dekumyoy, P., Komalamisra, C., et al. : Systematics of the subfamily Haplorchiinae (Trematoda: Heterophyidae), based on nuclear ribosomal DNA gene and ITS2 region. Parasitol Int 59(1):460-5, 2010.
13. Chuboon, S., Wongsawad, C. : Molecular identification of larval trematode in intermediate hosts from Chiang Mai, Thailand. Southeast Asian J Trop Med Publ Health 40(1):1216-20, 2009.
14. Chontananarth, T., Wongsawad, C. : *Haplorchis taichui* infection of the freshwater snails and molecular identification. Trend Res Sci & Tec 2(1):7-12, 2010.



15. Le, T.H., Donald, D.B., McManus, P. : Mitochondrial genomes of parasitic flatworms. *Trends in Parasitol.* 18(5):206-13, 2001.
16. Lee, S.U., Huh, S., Sohn, W., et al. : Sequence comparisons of 28S ribosomal DNA and mitochondrial cytochrome c oxidase subunit I of *Metagonimus yokogawai*, *M. takahashii* and *M. miyatai*. *Korean J Parasitol.* 42(3):129-35, 2004.
17. Brandt, R.A.M. : The non-marine aquatic mollusca of Thailand. *Arch Moll Band.* pp 27-340, 1974.
18. Schell, S.C. : How to know the trematode. W.M. C. Brown Company Publishers. America. pp 5-40, 1970.
19. Ito J. : Studies on cercariae in Japan. Shizuoka University. Japan, pp 80-150, 1980.
20. Skov, J., Kania, P.W., Dalsgaard, A., et al. : Life cycle stages of heterophyid trematode in Vietnamese freshwater fishes traced by molecular and morphometric Methods. *Vet Parasitol* (160):66-75, 2009.
21. Yu, J.R., Chung, J.S., Huh S., et al. : PCR-RFLP patterns of three kinds of *Metagonimus* in Korea. *Korean J Parasitol.* 35(4):271-6, 1997.
22. Mard-arhin, N., Prawang, T., Wongsawad, C. : Helminths of freshwater animals from five provinces in northern Thailand. *Southeast Asian J Trop Med Public Health* 32(2):206-14, 2001.
23. Wongsawad, C., Kumchoo, K. : Studies on prevalence and intensity of *Transversotrema patialensis* (Trematoda: Transversotrematidae) in the snail intermediate host, *Thiara scabra*. *J Med and Appl Malacol* 10(1):37-40, 2000.
24. Ukong, S., Krailas, D., Dangprasert, T. et al. : Studies on the morphology of cercariae obtained from freshwater snails at Erawan waterfall, Erawan national park, Thailand. *Southeast Asian J Trop Med Public Health* 38(2):302-12, 2007.
25. Radomyos, B. Wongsaroj, T., Wilairata, P., et al. : Opisthorchiasis and intestinal fluke infections in northern Thailand. *Southeast Asian J Trop Med Public Health* 29(1):123-7, 1988.
26. Nithikathkul, C., Wongsawad, C. : Prevalence of *Haplorchis taichui* and *Haplorchoides* sp. metacercariae in freshwater fish from water reservoirs, Chiang Mai, Thailand. *Korean J Parasitol.* 46(1):109-12, 2008.